

Keep coccidiosis under control - naturally!



By **Dr. Inge Heinzl**, Editor, **Madalina Diaconu**, Product Manager Pretect D, and **Dr. Ajay Awati**, Global Category Manager Gut Health & Nutrition, EW Nutrition

Often you have an extensive coccidiosis control program in place. You don't observe any clinical signs of coccidiosis. However, at the end of the cycle, you record significantly lower body weight and a higher FCR. There is a high probability that your animals have subclinical coccidiosis. This article digs deeper into understanding why birds don't perform as they should, why subclinical coccidiosis occurs on the farm, and why drug resistance is an important factor.

Subclinical coccidiosis - a silent enemy

Clinical coccidiosis is clearly characterized by severe diarrhea, high mortality rates, reduced feed/water intake, and weight loss. By contrast, subclinical Coccidiosis does not display any visual signs and often remains undetected.

According to De Gussem (2008), the damages caused by subclinical coccidiosis can reach up to 70% of the total cost of coccidiosis control treatments, ranging from US\$ 2.3 billion to US\$ 13.8 billion/year in 2020 worldwide (De Gussem, 2008; Ferreira da Cunha, 2020; Blake et al., 2020).

Monitoring coccidiosis occurrence on the farm

There are several tools available to evaluate the level of infection. The most common ones are:

Lesion scoring – is used to evaluate the damages caused by coccidiosis in the intestinal tract. Lesion scoring gives insight into the severity of the infection. Furthermore, based on the location of lesions in the GI tract, it is possible to determine the plausible *Eimeria* spp. responsible for the infection.

OPG (Oocyst per gram) – the number of oocysts per gram of feces indicates the level of shedding of oocysts in the manure, litter, and, eventually, in the farm environment. OPG levels may not give the exact severity of the infection in the bird but certainly provide a clear idea of its likely spread within the flock.

Ways to deal with coccidiosis on the farm

Different tools are widely used to prevent and treat coccidiosis:

Anticoccidials: Chemicals, ionophores

Vaccination: Natural strains, attenuated strains

Bio-shuttle: Vaccine + ionophore

Natural anticoccidials: Phytomolecules

These coccidiosis control programs are used depending on the farm history and the severity of the infection. Traditionally, treatment was heavily dependent on chemicals and ionophores. However, rampant and unbridled use of ionophores leads to resistance in *Eimeria* spp. on the farm, the failure of the control program, and significant performance losses, with high mortality due to coccidiosis. Therefore, the tools mentioned above are inserted in rotation or shuttle programs to minimize the generation of resistances. In a rotation program, the anticoccidial changes from flock to flock. In a shuttle program, the anticoccidial changes within one cycle according to the feed (Chapman, 1997).

However, this strategy is often not 100% effective due to a lack of diversity and overuse of certain tools within programs. The rigorous financial optimization of the program leads to the use of cost-effective but marginally effective solutions. These factors over the period weaken the program, which seems to work well but leads to resistance to anticoccidial drugs and sets up subclinical coccidiosis.

Resistances have been reported in the US (Jeffers, 1974, McDougald, 1981), South America (McDougald, 1987; Kawazoe and Di Fabio, 1994), Europe (Peeters et al., 1994; Bedrník et al., 1989; Stephan et al., 1997), Asia (Lan et al., 2017; Arabkhazaeli et al., 2013), and Africa (Ojimelukwe et al., 2018). Chapman and co-workers (1997) even stated that resistances were documented for all anticoccidial drugs employed at this time, and new products have not been approved for decades.

Resistance and subclinical coccidiosis can be approached naturally

When an anticoccidial has lost its effectiveness due to excessive use, some resistant coccidia survive. They can cause a mild course of the disease, subclinical coccidiosis, driving the costs high. Reducing the occurrence of resistance and subclinical coccidiosis can significantly decrease the expenses of coccidiosis control programs and, eventually, the cost of production.

Increasing consumer pressure to reduce the overall usage of drugs in animal production has driven innovation efforts to find natural solutions that can be effectively used within coccidiosis control programs.

However, this shift was not easy for the producers. Lack of reliable data, poor understanding of the mode of action, lack of quality optimization, and unsubstantiated claims led to the failure of many earlier-generation natural solutions.

However, the consumer-driven movement to find natural solutions to animal gut health issues has recently led to relentless innovation in this area. Knowledge, research, and technological developments are now ready to offer solutions that can be an effective part of the coccidia control program and open opportunities to make poultry production even more sustainable by reducing drug dependency.

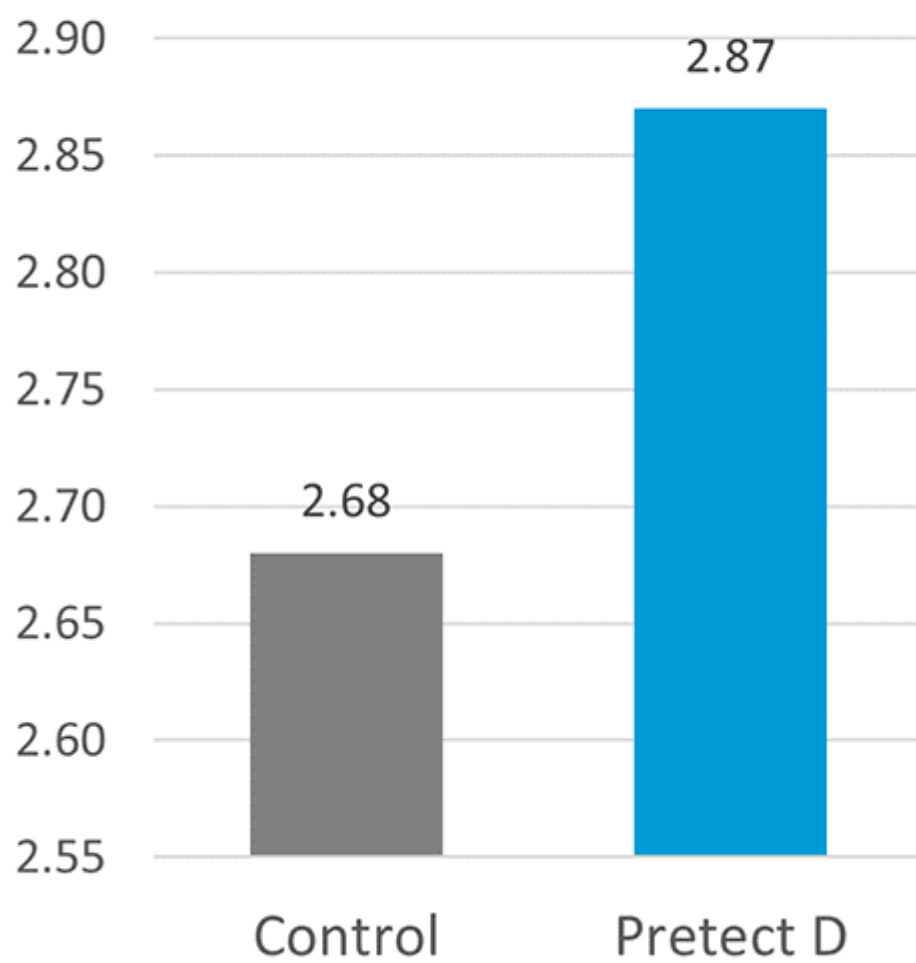
For centuries, phytochemicals have been used for their medicinal properties and effects on the health and well-being of animals and humans. In the case of coccidiosis, tannins and saponins have been proven to support animals in coping with this disease. Tannic acids and tannic acid extracts strengthen the intestinal barrier by reducing oxidative stress and inflammation (Tonda et al., 2018). On the other hand, saponins lessen the shedding of oocysts, improve the lesion score, and, in the case of an acute infection, the occurrence of bloody diarrhea (Youssef et al., 2021).

These natural substances can be integrated into shuttle or rotation programs to reduce the use of anticoccidials and, therefore, minimize resistance development.

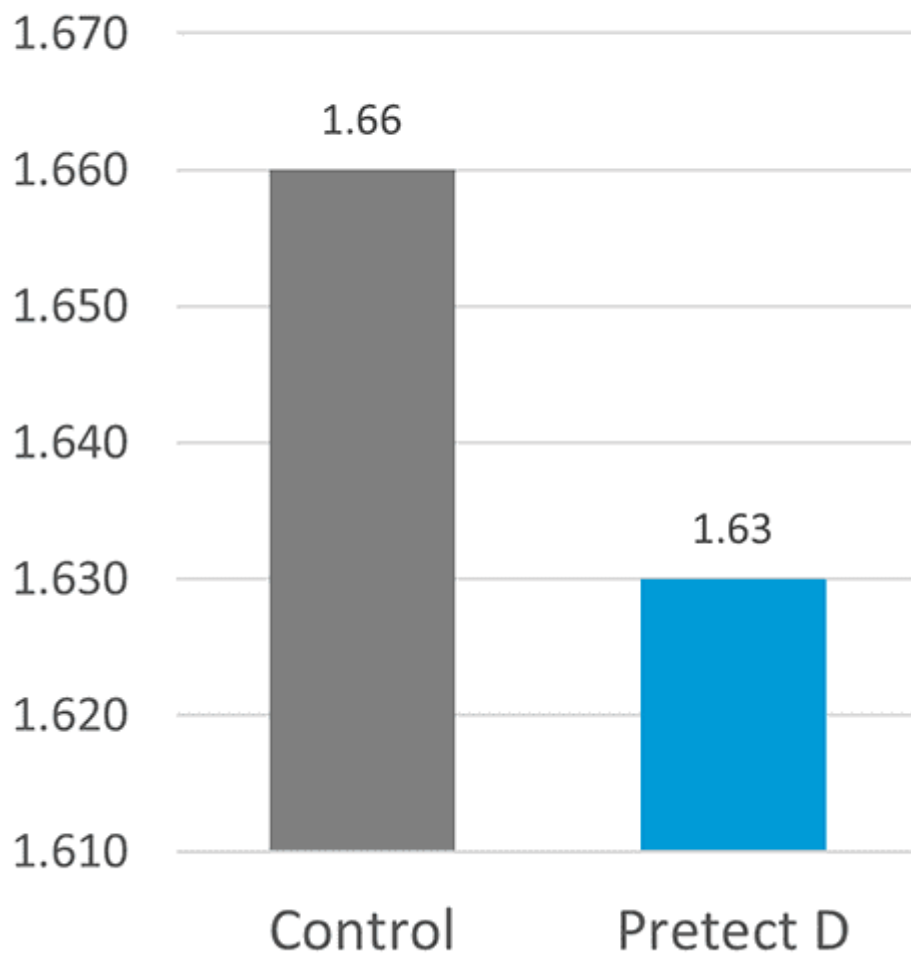
Pretect D: Coccidiosis programs can be strengthened naturally!

In an EU field trial conducted with more than 200 000 birds, [Prepect D](#) (a natural phytochemical-based product designed to increase the efficacy of coccidiosis control) was used in the shuttle program together with ionophores. The trial provided excellent results on zootechnical performance (figures 1-4).

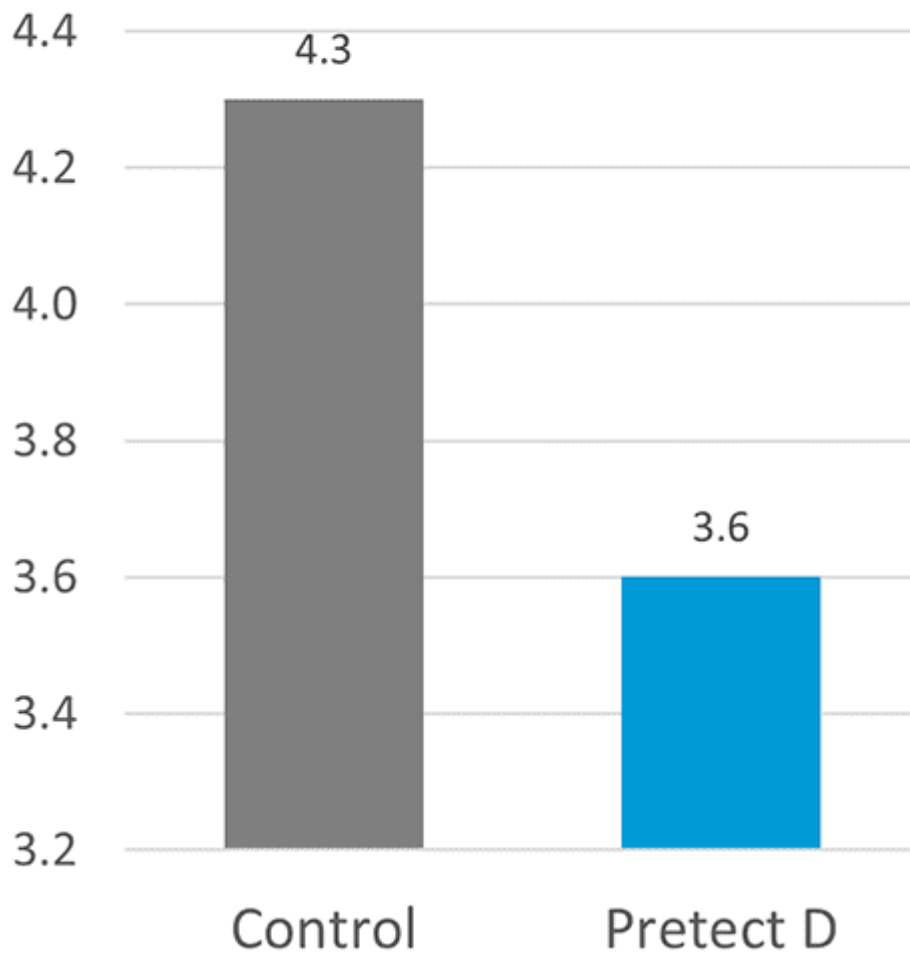
Bodyweight (kg)

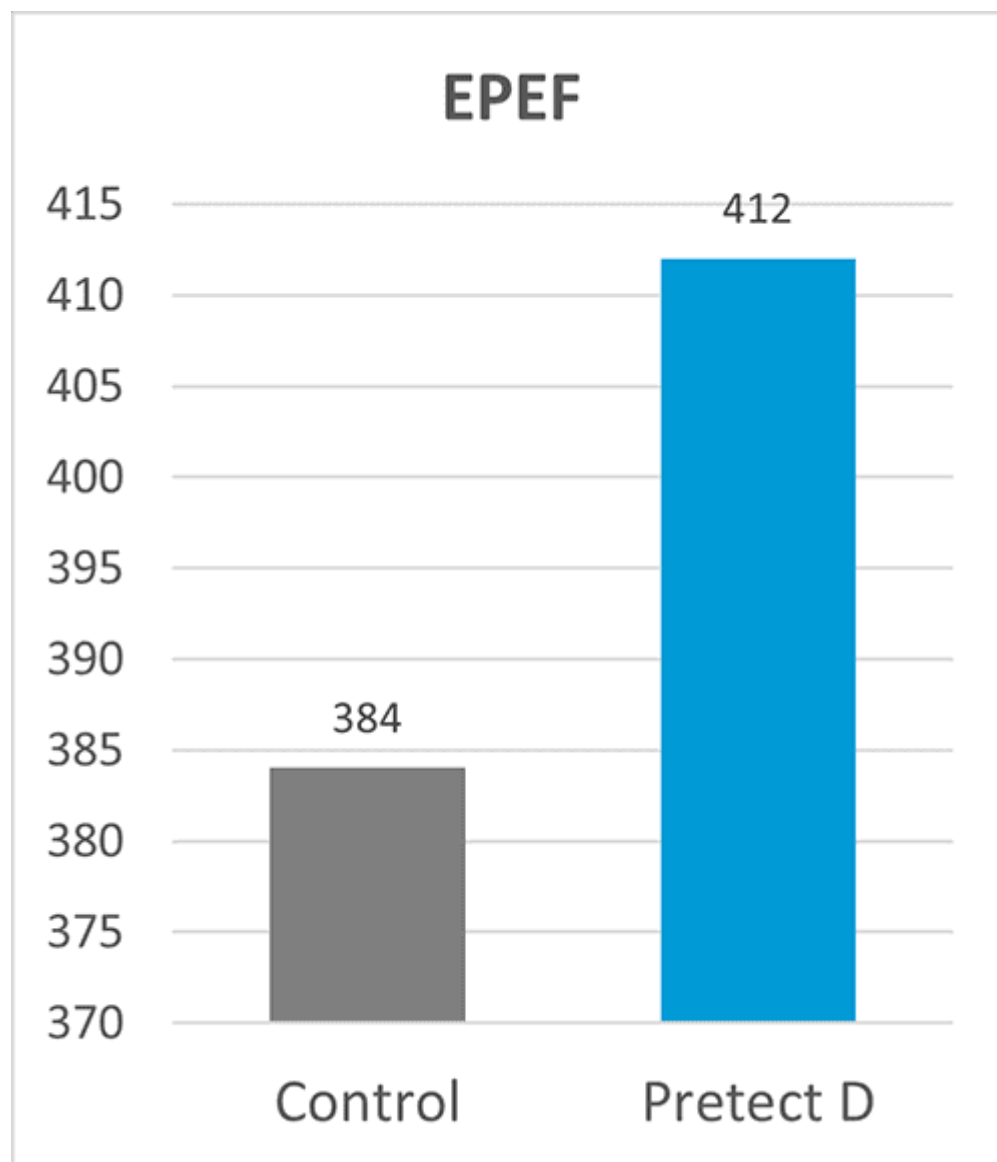


FCR



Mortality (%)





Figures 1-4: Zootechnical performance of broilers with Pretect D included in the shuttle program

Trials show that **Pretect D supports the efficiency of coccidiosis control programs** by impairing the *Eimeria* development cycle when used in combination with vaccines, ionophores, and chemicals as part of the shuttle or rotation program:

- It protects the epithelium from inflammatory and oxidative damage
- It promotes the restoration of the mucosal barrier function

Table 1 exemplifies one way of including a natural solution (Pretect D) in actual coccidiosis control programs.

	Week 1	Week 2	Week 3	Week 4	Week 5	Week 6 - Onwards
1	Coccidiostat					Prectect D
2	Coccidiostat			Prectect D		
3	Prectect D					
4	Vaccine			Prectect D		

Table 1: Example of including Pretect D into coccidiosis control programs

Natural solutions suit both farmers and consumers

With phytomolecules partly replacing anticoccidials in rotation or shuttle programs, the use of anticoccidials in poultry production can be decreased. On the one hand, this answers consumers' demand; on the other hand, it leads to a push-back of resistances in the long run. The returning effectiveness of the anticoccidials can reduce subclinical coccidiosis, leading to lower costs spent on this disease and a higher profit for the farmers.

References:

- Arabkhazaeli, F., M. Modrisanei, S. Nabian, B. Mansoori, and A. Madani. "Evaluating the Resistance of Eimeria spp. Field Isolates to Anticoccidial Drugs Using Three Different Indices." *Iran J Parasitol.* 8, no. 2 (2013): 234-41.
- Bedrník, P., P. Jurkovič, J. Kučera, and A. Firmanová. "Cross Resistance to the IONOPHOROUS Polyether Anticoccidial Drugs IN Eimeria Tenella Isolates from Czechoslovakia." *Poultry Science* 68, no. 1 (1989): 89-93. <https://doi.org/10.3382/ps.0680089>
- Blake, Damer P., Jolene Knox, Ben Dehaeck, Ben Huntington, Thilak Rathinam, Venu Ravipati, Simeon Ayoade, et al. "Re-Calculating the Cost of Coccidiosis in Chickens." *Veterinary Research* 51, no. 1 (2020). <https://doi.org/10.1186/s13567-020-00837-2>
- Chapman, H. D. "Biochemical, Genetic and Applied Aspects of Drug Resistance in Eimeria Parasites of the Fowl." *Avian Pathology* 26, no. 2 (1997): 221-44. <https://doi.org/10.1080/03079459708419208>.
- De Gussem, M., and S. Huang. "The Control of Coccidiosis in Poultry." *International Poultry Production* 16, no. 5 (2008): 7-9.
- Ferreira da Cunha, Anderson, Elizabeth Santin, and Michael Kogut. "Editorial: Poultry Coccidiosis: Strategies to Understand and Control." *Frontiers in Veterinary Science* 7 (2020). <https://doi.org/10.3389/fvets.2020.599322>
- Jeffers, T. K. "Eimeria Acervulina and E. Maxima: Incidence and Anticoccidial Drug Resistance of Isolants in Major Broiler-Producing Areas." *Avian Diseases* 18, no. 3 (1974): 331. <https://doi.org/10.2307/1589101>
- Kawazoe, Urara, and J. Di Fabio. "Resistance to DICLAZURIL in Field Isolates OfEimeriaspecies Obtained from Commercial BROILER Flocks in Brazil." *Avian Pathology* 23, no. 2 (1994): 305-11. <https://doi.org/10.1080/03079459408418998>
- Lan, L.-H., B.-B. Sun, B.-X.-Z. Zuo, X.-Q. Chen, and A.-F. Du. "Prevalence and Drug Resistance of Avian Eimeria Species in Broiler Chicken Farms of Zhejiang PROVINCE, CHINA." *Poultry Science* 96, no. 7 (2017): 2104-9. <https://doi.org/10.3382/ps.pew499>
- McDougald, L. R. "Anticoccidial Drug Resistance in the Southeastern United STATES: POLYETHER, IONOPHOROUS Drugs." *Avian Diseases* 25, no. 3 (1981): 600. <https://doi.org/10.2307/1589990>
- McDougald, Larry R., Jose Maria Silva, Juan Solis, and Mauricio Braga. "A Survey of Sensitivity to Anticoccidial Drugs in 60 Isolates of Coccidia from Broiler Chickens in Brazil and Argentina." *Avian Diseases* 31, no. 2 (1987): 287. <https://doi.org/10.2307/1590874>
- Ojimekwe, Agatha E., Deborah E. Emedhem, Gabriel O. Agu, Florence O. Nduka, and Austin E. Abah. "Populations of Eimeria Tenella Express Resistance to Commonly Used Anticoccidial Drugs in Southern Nigeria." *International Journal of Veterinary Science and Medicine* 6, no. 2 (2018): 192-200. <https://doi.org/10.1016/j.ijvsm.2018.06.003>
- Peeters, Johan E., Jef Derijcke, Mark Verlinden, and Ria Wyffels. "Sensitivity of AVIAN EIMERIA Spp. to Seven Chemical and Five Ionophore Anticoccidials in Five Belgian INTEGRATED Broiler Operations." *Avian Diseases* 38, no. 3 (1994): 483. <https://doi.org/10.2307/1592069>
- Stephan, B., M. Rommel, A. Dauschies, and A. Haberkorn. "Studies of Resistance to Anticoccidials IN Eimeria

Field Isolates and Pure Eimeria Strains." *Veterinary Parasitology* 69, no. 1-2 (1997): 19-29.
[https://doi.org/10.1016/s0304-4017\(96\)01096-5](https://doi.org/10.1016/s0304-4017(96)01096-5)

Tonda, RM, J.K. Rubach, B.S. Lumpkins, G.F. Mathis, and M.J. Poss. "Effects of Tannic Acid Extract on Performance and Intestinal Health of Broiler Chickens Following Coccidiosis Vaccination and/or a Mixed-Species Eimeria Challenge." *Poultry Science* 97, no. 9 (2018): 3031-42. <https://doi.org/10.3382/ps/pey158>

Youssef, Ibrahim M., Klaus Männer, and Jürgen Zentek. "Effect of Essential Oils or Saponins Alone or in Combination on Productive Performance, Intestinal Morphology and Digestive Enzymes' Activity of Broiler Chickens." *Journal of Animal Physiology and Animal Nutrition* 105, no. 1 (2020): 99-107.
<https://doi.org/10.1111/jpn.13431>

Necrotic enteritis: The complete overview



by **Inge Heinzl** and **Technical Team**, EW Nutrition

Necrotic enteritis is a profit killer in poultry production

Necrotic enteritis is the cause of USD 6 billion losses every year in global poultry production, corresponding to USD 0.0625 per bird (Wade and Keyburn, 2015). This controllable disease is on the rise. One reason is the voluntary or legally required reduction of antibiotics in animal production due to the increasing occurrence of antimicrobial resistance but also consumer demand. Another reason is the administration of live Coccidiosis vaccines and partial reduction of ionophores, which also show efficacy against Gram-positive bacteria (Williams, 2005).

Necrotic enteritis and coccidiosis are the most significant health problem in broilers (Hofacre et al., 2018).

The disease generally occurs in broiler chickens of 2-6 weeks of age. It is caused by an overgrowth of *Clostridium perfringens* type A and, to a lesser extent, type C in the small intestine. The toxins produced by *C. perfringens* also damage the intestinal wall.

Clinical and subclinical forms of NE - which one causes more significant losses?

The clinical form is obvious



Intestine showing signs of NE

...is characterized by acute, dark diarrhea resulting in wet litter and suddenly increasing flock mortality of up to 1% per day after the first clinical signs appear (Ducatelle and Van Immerseel, 2010), sometimes summing up to mortality rates of 50% (Van der Sluis, 2013). The birds have ruffled feathers, lethargy, and inappetence.

Necropsy typically shows ballooned small intestines with a roughened appearing mucosal surface, lesions, and brownish (diphtheritic) pseudomembranes. There is a lot of watery brown, blood-tinged fluid and a foul odor during post-mortem examination. The liver is dark, swollen, and firm, and the gall bladder is distended (Hofacre et al., 2018).

In the case of **peracute** Necrotic Enteritis, birds may die without showing any preliminary signs.

The subclinical form often only can be noticed at the end of the cycle

When birds suffer from the subclinical form, chronic damage to the intestinal mucosa and an increased quantity of mucus in the small intestine lead to impaired digestion and absorption of nutrients resulting in poor growth performance. The deteriorated feed conversion and the resulting decreased performance become noticeable around day 35 of age. As feed contributes approximately 65-75% of the input cost to produce a broiler chicken, poor feed conversion increases production costs and significantly influences profitability. Often, due to a lack of clear symptoms, this subclinical disease remains untreated and

permanently impacts the efficiency of production.



The pathogen causing NE - a ubiquitous bacterium

Responsible for Necrotic Enteritis are Gram-positive, anaerobic bacteria, specific strains of *Clostridium perfringens* type A and, to a lesser extent, type C (Keyburn et al., 2008).



Clostridia primarily occur in the soil where organic substances are degraded, in sewage, and in the gastrointestinal tract of animals and humans. These bacteria produce spores, which are extremely resistant to environmental impact (heat, irradiation, exsiccation), some disinfectants, and can survive for several years. Under suitable conditions, *C. perfringens* spores can even proliferate in feed or litter.

Clostridium perfringens is a “natural inhabitant” of the intestine of chickens. In healthy birds, it occurs in a mixture of diverse strains at 10^2 - 10^4 CFU/g of digesta (McDevitt et al., 2006). The disease starts when *C. perfringens* proliferates in the small intestine, usually due to a combination of factors such as high amount

protein, low immunity, and an imbalance in the gut flora. Then, the number rises to 10^7 - 10^9 CFU/g of digesta (Dahiya et al., 2005).

Highly important: NetB, a pore-forming toxin is a key virulence factor for NE

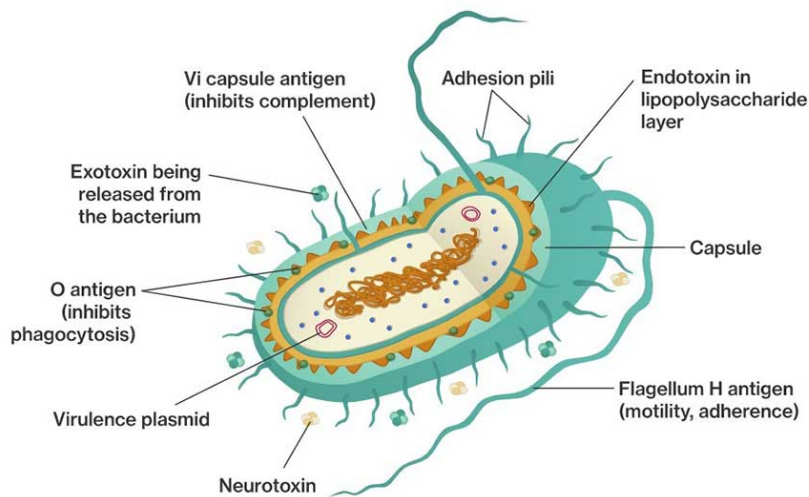
To establish in the host, *Clostridium* Spp. and other pathogens depend on virulence factors (see infobox). These virulence factors include for example “tools” for attachment, evasion or suppression of the host’s immune system, “tools” for getting nutrients, and “tools” for entry into intestinal cells. Over the years, the α -toxin produced by *C. perfringens* was assumed to be involved in the development of the disease and a key virulence factor. In 2008, Keyburn and coworkers found another key virulence factor by using a *C. perfringens* mutant unable to produce α -toxin, while still causing Necrotic Enteritis.

Thus, another toxin was identified occurring only in chickens suffering from Necrotic Enteritis: *C. perfringens* necrotic enteritis B-like toxin (NetB). NetB is a pore-forming toxin. Pore-forming toxins are exotoxins usually produced by pathogenic bacteria but may also be produced by other microorganisms. These toxins destroy the integrity of gut wall cell membranes. The leaking cell contents serve as nutrients for the bacteria. If immune cells are destroyed, an immune reaction might be partially impaired.

Additionally, pathogenic strains of *C. perfringens* produce bacteriocins – the most important is Perfrin (Timbermont et al., 2014) – to inhibit the proliferation of harmless *Clostridium* Spp. strains and to replace the normal intestinal flora of chickens (Riaz et al., 2017).

Examples of virulence factors

1. **Adhesins**
Enable the pathogen to adhere or attach within the target host site, e.g. via fimbria. Pili enable the exchange of RNA or DNA between pathogens.
2. **Invasion factors**
Facilitate the penetration and the distribution of the pathogens in the host tissue (invasion and spreading enzymes). For example: hyaluronidase attacking the hyaluronic acid of the connective tissue or flagella enabling the pathogens to actively move.
3. **Toxins**
Damage the function of the host cells or destroy them (e.g. endotoxins – lipopolysaccharides, exotoxins)
4. **Strategies of evasion**
Enable the pathogen to undergo the strategies of defense of the immune system (e.g. antiphagocytosis factors provide protection against an attack by phagocytes; specific antibodies are inactivated by enzymes).

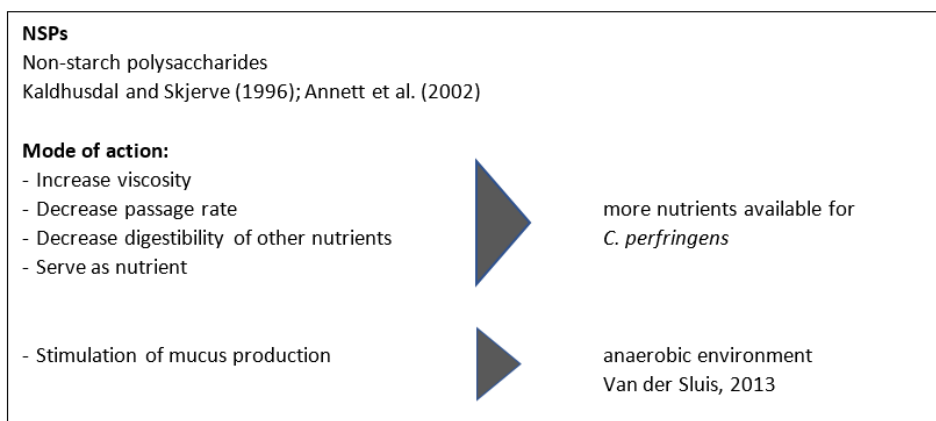


Predisposing factors favor the development of NE

A chicken with **optimal gut health** may be less **susceptible to NE**. Additional **predisposing factors** are necessary to allocate nutrients and make the gut environment suitable for the proliferation of these pathogens, enabling them to cause disease (Van Immerseel et al., 2008; Williams, 2005).

1. FEED: composition and particle size are critical

Feed plays a role in the development of Necrotic Enteritis that should not be underestimated. Here, substances creating an intestinal environment favorable for *C. perfringens* must be mentioned.



Protein and fat

Excessive content of protein

Poorly digestible protein

especially protein (fish meal) with high
Concentrations of zinc/glycine

Mode of action:

- High protein levels in the GIT
- Act as substrate for the bacteria

shift in balance of intestinal flora
(Antonissen et al., 2016)

Animal fat

Mode of action:

- Higher pH and higher bile salt conc.
- Promotion of coccis

increased growth of *C. perfringens*
(Knarreborg et al., 2002)

Particle size

Not uniform particle size

increased rates of NE
(Branton et al. 1987)

Finely-ground

Mode of action:

- gizzard musculature ↓
- pH ↑, GIT transit time ↓

proliferation of pathogenic microbes
(*Salmonella*, *Campylobacter*,
Clostridium...) ↓
(In comparison to coarse corn)
(Rougière and Carré, 2010 ; Santos et al., 2008;
Singh et al. 2014)

2. Mycotoxins create ideal conditions for NE

Mycotoxins harm gut integrity and create ideal conditions for the proliferation of *Clostridium perfringens*:

Mycotoxins do not have a direct effect on *C. perfringens* proliferation, toxin production, or NetB transcription. However, mycotoxins disrupt gut health integrity, creating a favorable environment for the pathogen. For example:

- DON provides good conditions for proliferation of *perfringens* by disrupting the intestinal barrier and damaging the epithelium. The possibly resulting permeability of the epithelium and a decreased absorption of dietary proteins can lead to a higher amount of proteins in the small intestine. These proteins may serve as nutrients for the pathogen (Antonissen et al., 2014).
- DON and other mycotoxins decrease the number of lactic acid producing bacteria indicating a shift in the microbial balance (Antonissen et al., 2016.)

3. Eimeria spp.: forming a perfect team with Clostridium perfringens

An intact intestinal epithelium is the best defense against potential pathogens such as *C. perfringens*. Here, coccidiosis comes into play. Moore (2016) showed that by damaging the gut epithelium, *Eimeria* species give *C. perfringens* access to the intestinal basal domains of the mucosal epithelium. Then, the first phase of the pathological process takes place and from there, *C. perfringens* invades the lamina propria. Damage to the epithelium follows (Olkowski et al., 2008). The plasma proteins leaking to the gut and the mucus produced are rich nutrient sources (Van Immerseel et al., 2004; Collier et al., 2008). A further impact of coccidiosis is shifting the microbial balance in the gut by decreasing the number of e.g.,

Candidatus savagella which activates the innate immune defense.

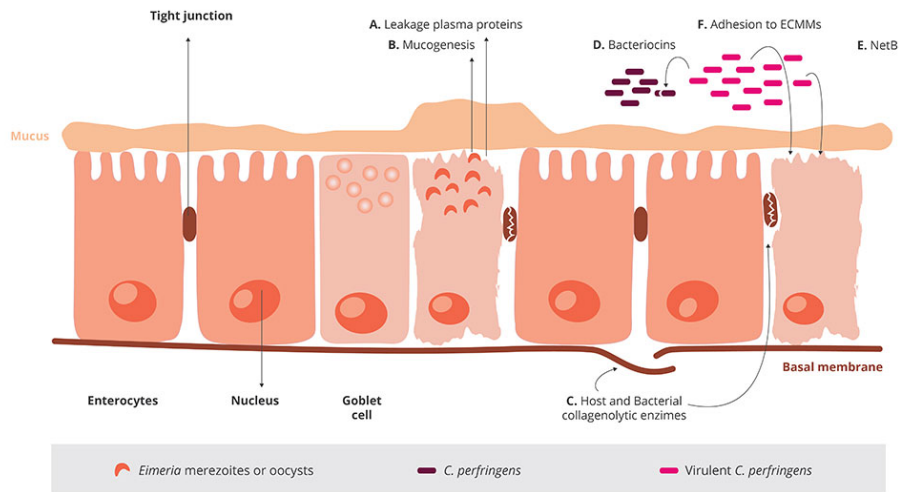


Figure 1

Figure 1:

- A. *Eimeria* induce leakage of plasma proteins by killing epithelial cells
- B. They enhance mucus production in the intestine

A+B lead to an increase in available nutrients and create an environment favorable for the proliferation of *perfringens*

Not only *Eimeria* Spp., also other pathogens (e.g. *Salmonella* Spp., Ascarid larvae, viruses) and agents, such as mycotoxins damaging the intestinal mucosa can pave the way for a *C. perfringens* infection. Predisposing factors like wet litter, the moisture of which is essential for the sporulation of *Eimeria* Spp. oocysts, must also be considered as promoting factors for Necrotic Enteritis (Williams, 2005).

4. Immunosuppressive Factors: Bacteria, viruses..., and stress

Any factor which induces stress in the animals disrupts the balance of the intestinal flora. The resulting suppression of the immune system contributes to the risk of Necrotic Enteritis (Tsiouris, 2016).

Bacteria

Shivaramaiah and coworkers (2011) investigated a neonatal *Salmonella typhimurium* infection as a predisposing factor for NE. The early infection causes significant damage to the gut (Porter et al., 1998). Additionally, Hassan et al. (1994) showed that the challenge with *Salmonella typhimurium* negatively impacted the development of lymphocytes which might also promote a colonization of *Clostridium perfringens*.

Viruses

Infectious Bursal Disease is known to increase the severity of infections with salmonella, staphylococci, but also clostridia. Another clostridia-promoting viral disease is Marek's Disease.

Stress:

The intestinal tract is particularly sensitive to any type of stress. This stress can be caused by e.g. too high temperatures, high stocking densities, an abrupt change of feed.

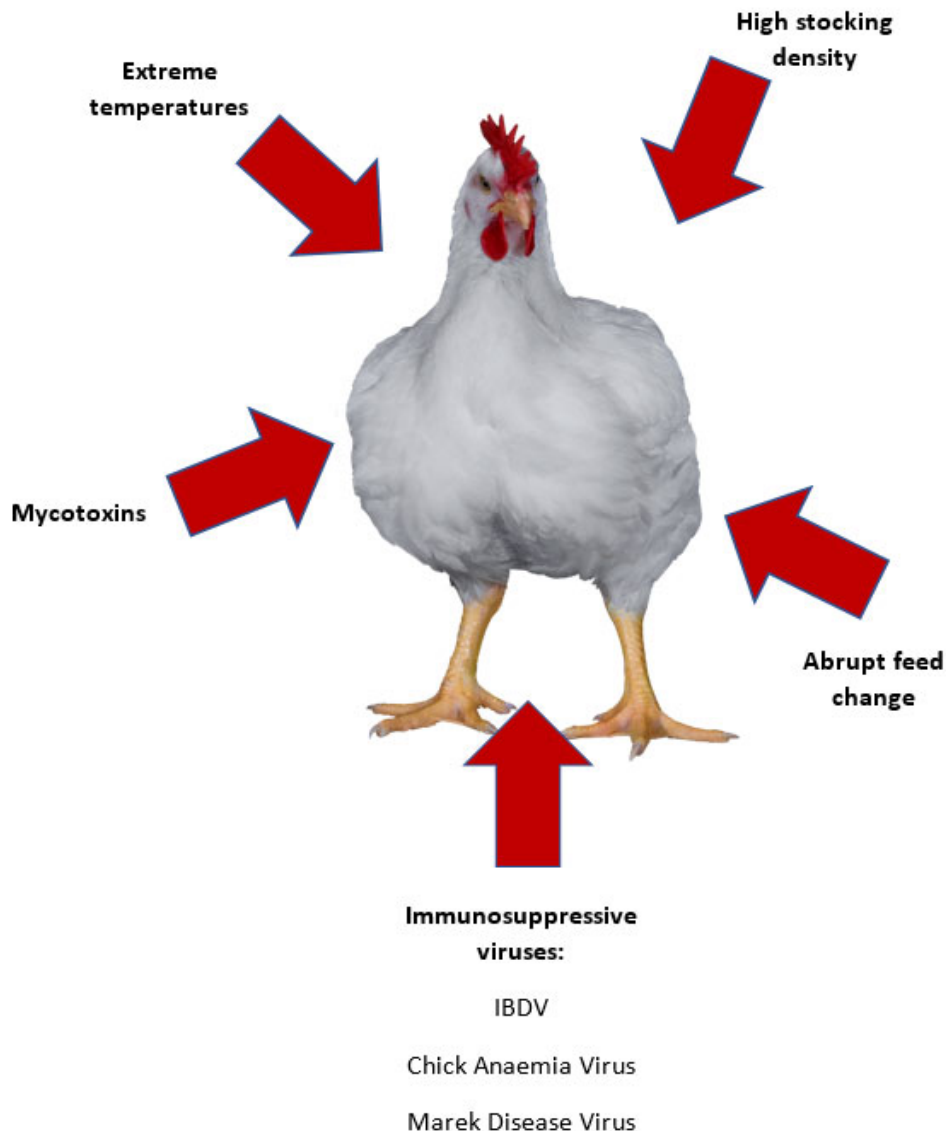


Figure 2: Predisposing factors weakening the birds and enabling Clostridium to attack

Treatment is necessary in the case of acute disease

In this instance, the farmer is obligated to consult a veterinarian and treat his birds.

It must be mentioned that, as the treatment takes place via feed or water, only birds which still consume water or feed may be treated.

Antibiotics are effective but also take a risk

Antibiotics targeting Gram-positive bacteria are commonly used for the treatment of acute NE. The antibiotic choice shall be addressed by a veterinarian, taking into account the mode of action and the presence of resistance genes in the farm/flock.

The profilactic use of antibiotics is not recommended and many countries have already [banned it in order to reduce antimicrobial resistance \(AMR\)](#).

Antimicrobial Resistance (AMR)

Some bacteria are less sensitive to certain antibiotics due to genetic mutations. They are able to:

- stimulate the production of enzymes, which break down or modify the antibiotics and inactivate them (1).
- eliminate entrances for antibiotics or promote the development of pumps, which discharge the antibiotic before taking effect (2).
- change or eliminate molecules to which the antibiotic would bind (targets for the antibiotics).

This means that, when the corresponding antibiotics are used, bacteria resistant against these antibiotics survive. Due to the fact that their competitors have been eliminated they are able to reproduce better.

Additionally, this resistance may be transferred by means of “resistance genes”

- to daughter cells
- via their intake from dead bacteria (3)
- through horizontal gene transfer (4)
- through viruses (5)

Every application of antibiotics promotes the development of resistance (Robert Koch Institute, 2019). A short-term use, [better biosecurity](#), or an application at low dosage give the bacteria a better chance to adapt.

Bacteriophages would be possible but are still disputed

Experimental use of phage treatments has shown to be effective in reducing disease progression and symptoms of Necrotic Enteritis (Miller et al., 2010). By oral application of a bacteriophage cocktail, Miller and coworkers could reduce mortality by 92% in *C. perfringens*-challenged broilers compared to the untreated control.

Mode of action: the endolysins, highly evolved enzymes produced by bacteriophages, are able to digest the bacterial cell wall for phage progeny release (Fischetti, 2010). However, phages are still not approved by the EFSA.

Excurs:

Antimicrobial Resistance (AMR) Some bacteria are less sensitive to certain antibiotics due to genetic mutations. They are able to:

- stimulate the production of enzymes, which break down or modify the antibiotics and inactivate them (1).
- eliminate entrances for antibiotics or promote the development of pumps, which discharge the antibiotic before taking effect (2).
- change or eliminate molecules to which the antibiotic would bind (targets for the antibiotics) (figure 3)

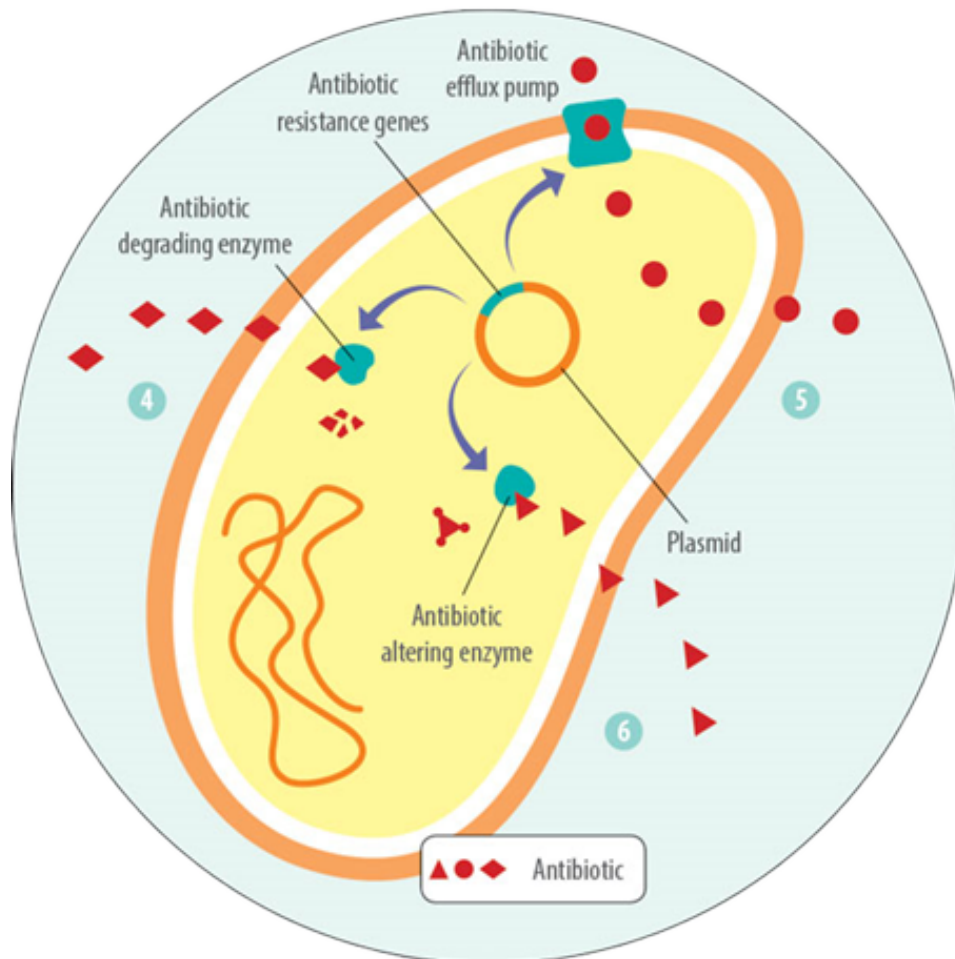


Figure 3: Possibilities of a bacterial cell to defend itself against antibiotics

This means that, when the corresponding antibiotics are used, bacteria resistant against these antibiotics survive. Due to the fact that their competitors have been eliminated they are able to reproduce better. Additionally, this resistance may be transferred by means of “resistance genes”

- to daughter cells
- via their intake from dead bacteria (3)
- through horizontal gene transfer (4)
- through viruses (5) (figure 4)

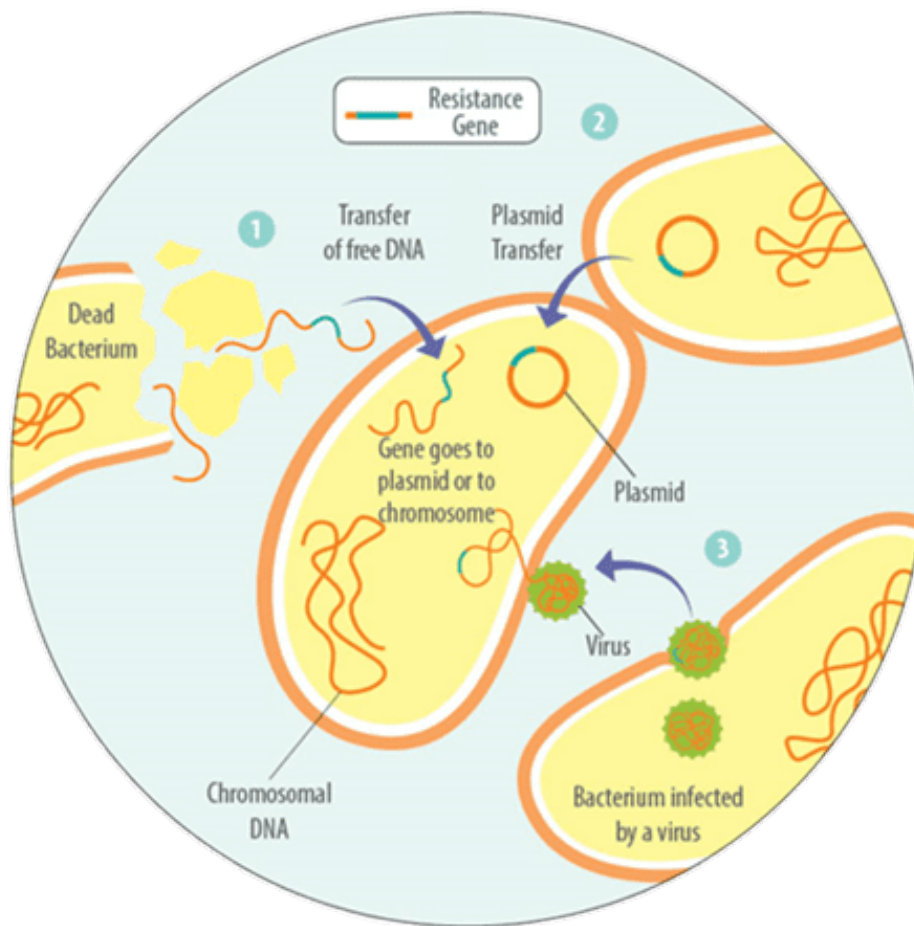


Figure 4: Possibilities to transfer resistance to other bacterial cells

Every application of antibiotics promotes the development of resistance (Robert Koch Institute, 2019). A short-term use or an application at low dosage give the bacteria a better chance to adapt.

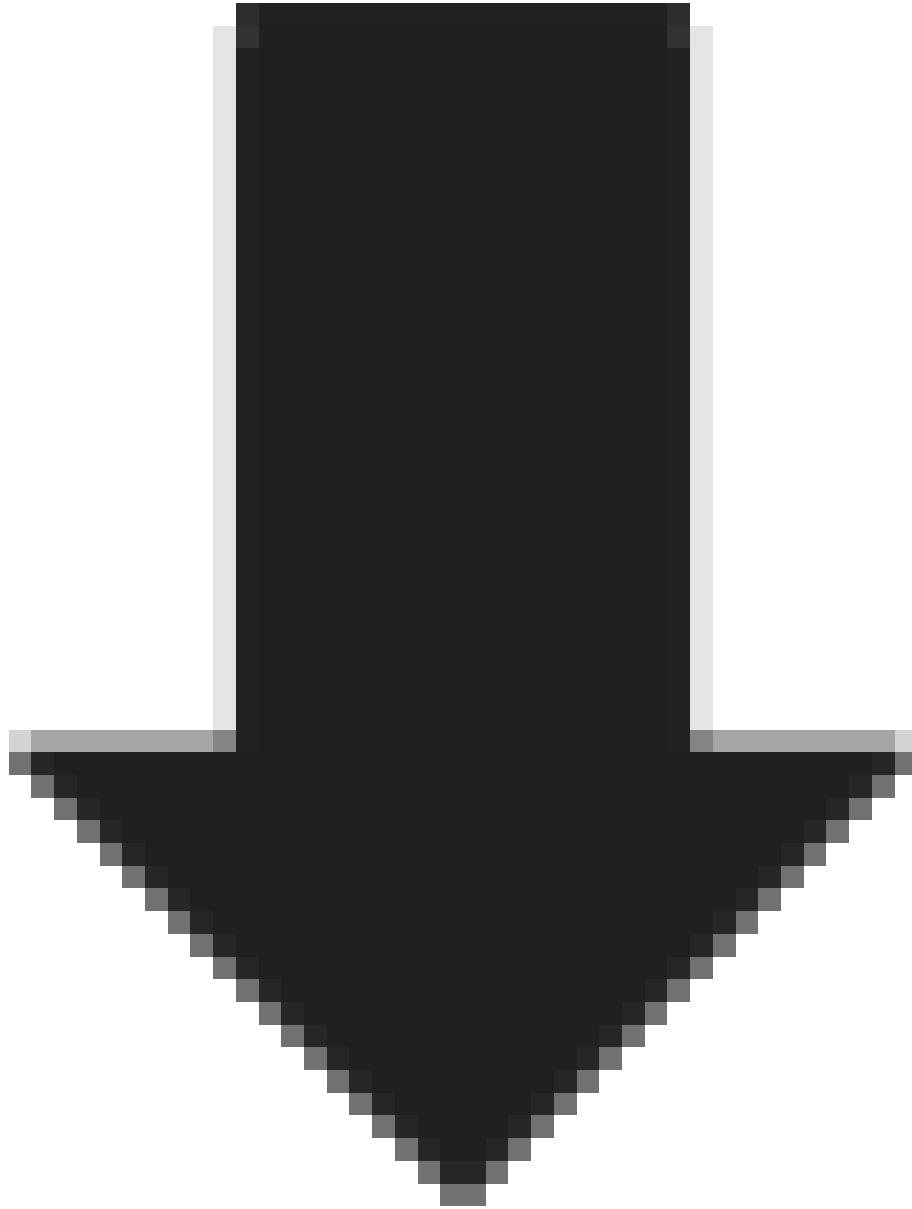
Preventing a disease is always better than its treatment!

But how to do it? Preventing the conditions that favor the proliferation of *Clostridium perfringens* and strengthening the host's immune response lowers the probability of disease. Besides eliminating the predisposing factors, the main targets are:

- **Balance of the gut flora**
- **Optimization of gut function and integrity**
- **Maintenance of immunity**

1. Biosecurity is of the highest importance!

There is evidence that most *Clostridium* strains isolated from birds suffering from Necrotic Enteritis could induce the disease experimentally, while strains isolated from healthy birds cannot. This confirms that only specific strains are problematic (Ducatelle and Van Immerseel, 2010). So, it's of highest importance to avoid introducing these pathogenic strains to the farm.



Strict biosecurity measures!

Separate clothing, boots, and hand washing/disinfecting facilities in each poultry house

More than 14 days of down time between flocks

2. Specific measures against coccidiosis

1. Vaccination

According to parasitologists, 7 to 9 *Eimeria* species are found in chickens, and they do not cross-protect against each other. An effective vaccination must contain sporulated oocysts of the most critical pathogenic *Eimeria* species (*E. acervulina*, *E. maxima*, *E. tenella*, *E. necatrix*, and *E. brunetti*). The more species contained in the vaccine, the better. However, if not applied the correct way, vaccines can be ineffective or cause reactions in the birds that might lead to NE (Mitchell, 2017).

2. Anticoccidials

Alternate use of chemicals (synthetic compounds) and ionophores (polyether antibiotics) with different modes of action is important to avoid development of resistance.

Ionophores have a specific mode of action and kill oocysts before they are able to infect birds. Being very small, ionophore molecules can be taken up and diffused into the outer membrane of the sporozoite. There, it decreases the concentration gradient leading to an accumulation of water within the sporozoite causing its bursting.

3. Diet - favorable for the birds but not for clostridium!

Minimizing non-starch polysaccharides (NSPs) in cereals



To prevent a “feeding” of *Clostridium perfringens*, high content of water-soluble but indigestible NSPs such as wheat, wheat by-products, and barley should be avoided or at least minimized. Additionally, xylanases should be included in the feed formulation to reduce the deleterious effects of NSPs and improve feed energy utilization. Instead of these cereals, maize could be included in the diet. It is considered a perfect ingredient in broiler diets due to its high energy content and high nutrient availability.

Formulating low protein diets/diets with highly digestible amino acids

Feeding low-protein diets supplemented with crystalline amino acids might be beneficial to reduce the risk of Necrotic Enteritis (Dahiya et al., 2007). To improve protein digestibility and therefore reduce the proliferation of *C. perfringens*, proteases may be added to the feed.

Avoiding/Minimizing animal fats in the diet

Animal fats tend to increase the counts of *Clostridium perfringens*; thus, they should be replaced by vegetable fat sources.

Feed form is decisive

In terms of feed form, Engberg et al. (2002) found that birds fed pellets showed a reduced number of *Clostridium perfringens* in the caeca and the rectum than mash-fed birds. Branton and coworkers (1987) reported a lower mortality by feeding roller-milled (coarsely ground) than hammer-milled feed.

4. Additives

Additives can be used either to prevent the proliferation of *Clostridium perfringens* or to change the environmental conditions in a way that proliferation of *C. perfringens* is prevented.

1. Probiotics directly support the balance of the microbiome

These live microbial supplements can be used to help to establish, maintain or re-establish the intestinal microflora.

Mode of action:

They compete with pathogenic bacteria for substrates and attachment sites and produce antimicrobial substances inhibiting the growth of pathogenic bacteria (Gillor et al., 2008). They bind and neutralize enterotoxins (Mathipa and Thantsha, 2017) and promote immune function of the host (Yang et al., 2012)

2. Prebiotics indirectly promote the microbiome

These feed ingredients serve as substrates to promote beneficial bacteria in the intestine.

Mode of action:

D-mannose or fructose, starches non-digestible by birds, selectively stimulate the growth and the activity of the “good” gut flora. Fructooligosaccharides decrease *C. perfringens* and *E. coli* in the gut and increase the diversity of *Lactobacillus* Spp. (Kim et al., 2011). Galactooligosaccharides, in combination with a *B. lactis*-based probiotic, have been reported to selectively promote the proliferation of *Bifidobacterium* Spp. (Jung et al., 2008).

3. Organic acids support gut health

[Organic acids](#) are often used in animal diets to improve intestinal health.

Mode of action:

A decreased pH promotes beneficial bacteria. Caprylic acid suppresses *C. perfringens* but also *Salmonella* spp. by inhibiting their utilization of glucose (Skrivanova et al., 2006). Lauric, citric, oleic, and linoleic acid, as well as medium-chain fatty acids (C8-C14), impede the growth of *C. perfringens*.

A trial with different organic acid products showed high efficacy for Acidomix AFG and Acidomix AFL against *Clostridium perfringens* as well as against *Salmonella enterica*. For the test, 50 µl solution containing different microorganisms (reference strains of *S. enterica* and *C. perfringens*; conc. 10⁵ CFU/ml) together with 50 µl of increasing concentrations of various organic acids/organic acid products (Acidomix) were pipetted into microdilution plates. After the respective incubation, the MICs of every organic acid/organic acid product were calculated.

Figure 5 shows the minimum inhibiting concentrations (MIC). For Acidomix AFL and AFG, lower concentrations than for fumaric, lactic, and propionic acid were needed to inhibit the growth of *Salmonella enterica* and *Clostridium perfringens*.

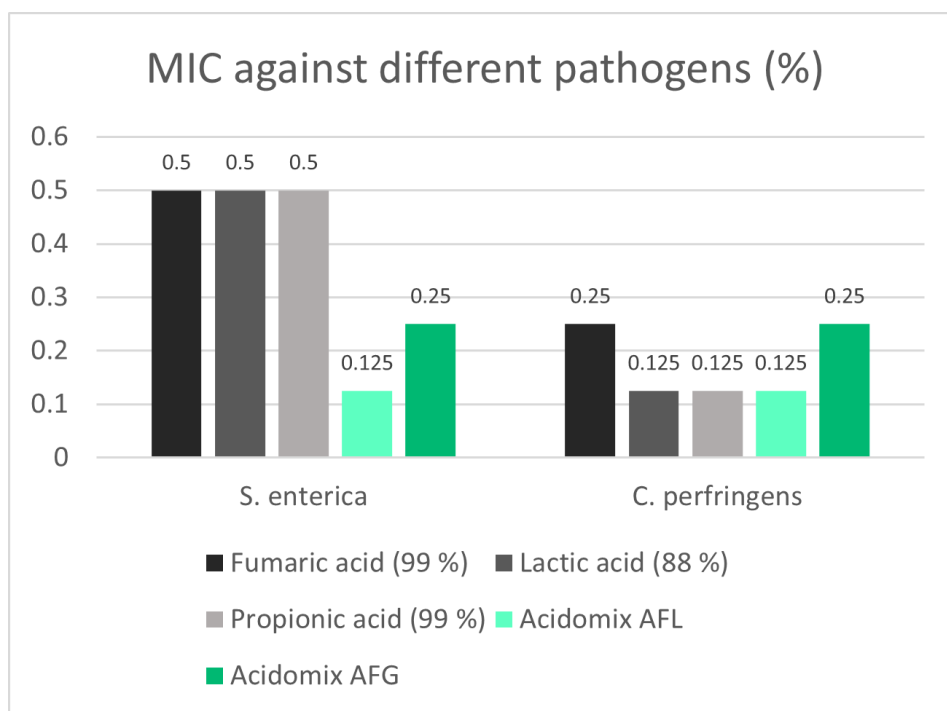


Figure 5: Minimal inhibiting concentrations of Acidomix AFL and Acidomix AFG against *Salmonella enterica* and *Clostridium perfringens*

Phytomolecules : different types are available against NE

[Phytomolecules](#), also known as secondary plant compounds, have been used against pathogens for centuries. In general, two subgroups of these substances are known as effective against *Clostridium perfringens*: Tannins and Essential Oils.

Tannins

Many studies have shown the efficacy of [tannins](#) against different pathogens such as helminths, *Eimeria* spp., viruses, and bacteria. Extracts from the chestnut and quebracho trees are effective not only against *C. perfringens* but also its toxins (Elizando et al., 2010). Tannins act against *Eimeria* spp. (Cejas et al., 2011) and *Salmonella* Sp., two predisposing factors for NE.

A trial was conducted with Pretect D, a product based on tannins and saponins, to show its efficacy against coccidia, one of the predisposing factors of NE. For the 35-day study conducted at a commercial research facility in the US, 1800 one-day-old Cobb 500 broilers were divided into four groups of 450 birds each (with 9 replicates & 50 birds per replicate). They all received the standard feed of the farm (Starter D0-D21, Grower D22-D35).

The challenge was given in the form of a freshly prepared mixed inoculum with *E. acervulina* (100,000 oocysts/ bird), *E. maxima* (50,000 oocysts/ bird), and *E. tenella* (75,000 oocysts/ bird). The inoculum was mixed into the feed in the base of each pen's tube feeder.

The oocyst count per gram of feces (OPG) was done on D21, D27 & D35. The cocci lesion scoring (CLS) was done on D27 following Johnson and Reid (1970) with 0=normal; 4=most

Group	Challenge	Additive
Non-challenged Control (NC)	No	No
Challenged Control (CC)	Yes	No
CC + Ionophore	Yes	Ionophore@60ppm
CC + Pretect D	Yes	Pretect D@500ppm

The trial showed that, due to Pretect D, the lesion score showed a lower value indicating that lesions could be reduced or were less severe, which can be seen in figure 6:

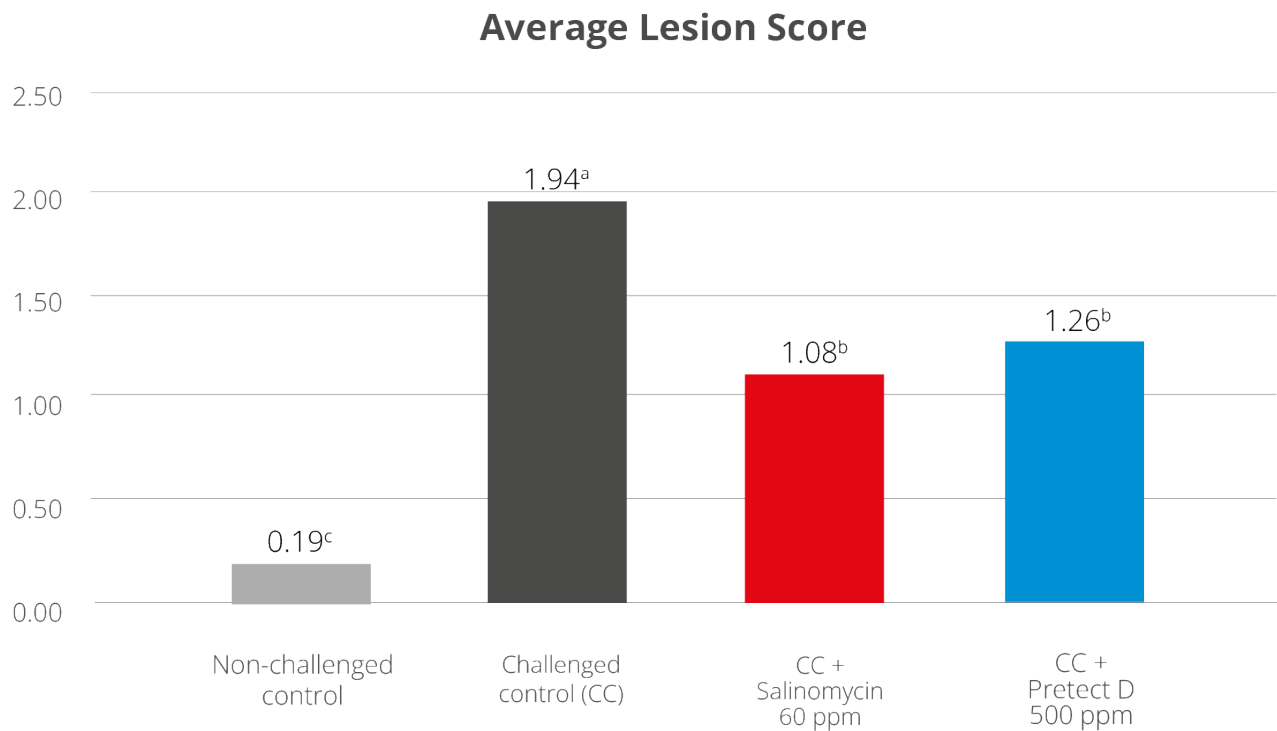


Figure 6: Average lesion score

Essential Oils

Their hydrophobic characteristic enables them to interact with the lipids of the membrane of *C. perfringens*. They can incorporate into the bacterial membrane and disrupt its integrity, increasing the permeability of the cell membrane for ions and other small molecules such as ATP and leading to the decrease of the electrochemical gradient above the cell membrane and the loss of the cell's energy equivalents. Besides their direct effect on *Clostridium spp.*, many phytochemicals improve gut health and help prevent the proliferation of *Clostridium spp.* And, therefore, Necrotic Enteritis.

An In vitro-trial shows Ventar D reducing *Clostridia* and sparing the beneficial lactobacilli. In this trial, the bacteria (*Clostridium perfringens*, *Lactobacillus agilis* S73, and *Lactobacillus plantarum*) were cultured under favorable conditions (RCM, 37°C, anaerobe for *Clostridium perfringens*, and MRS, 37°C, 5 % CO₂ for Lactobacilli) and exposed to different concentrations of Ventar D (0 µg/ml – control, 500 µg/ml, 750 µg/ml, and 1000 µg/ml).

The results of the trial with *Clostridium perfringens* are shown in figure 7.

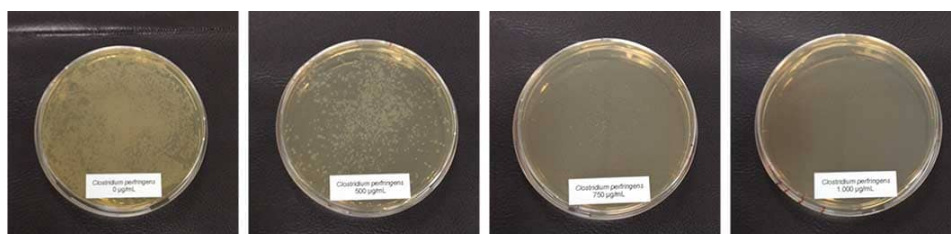


Figure 7: Different concentrations of Ventar D added to *Clostridium perfringens* cultures

Here, a significant reduction of colonies could already be observed at a concentration of 500 µg/ml of Ventar D. With 750 µg/ml, only a few colonies remained, and at a Ventar D concentration of 1000 µg/ml, *Clostridium perfringens* didn't grow anymore.

In contrast, the Lactobacilli showed a different picture: only at the higher concentration (1250 µg/ml of Ventar D) did *Lactobacillus plantarum* and *Lactobacillus agilis* S73 show a slight growth reduction (figure 8).

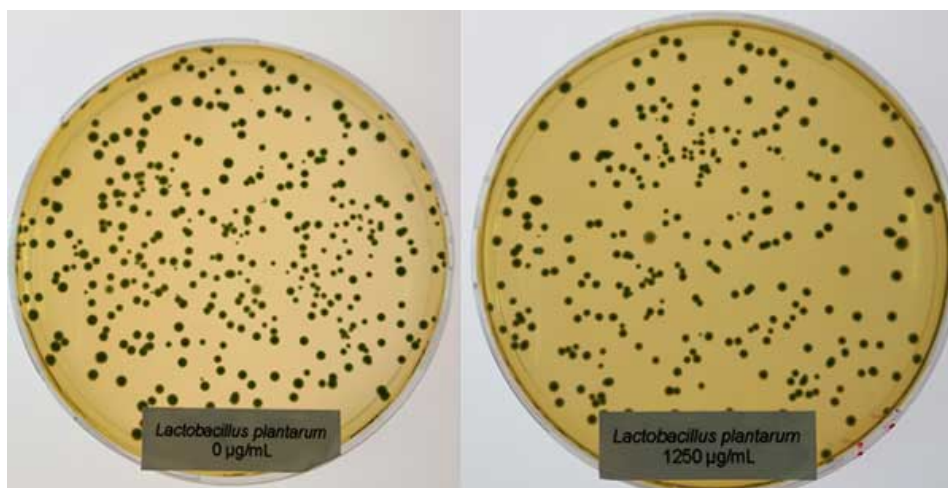


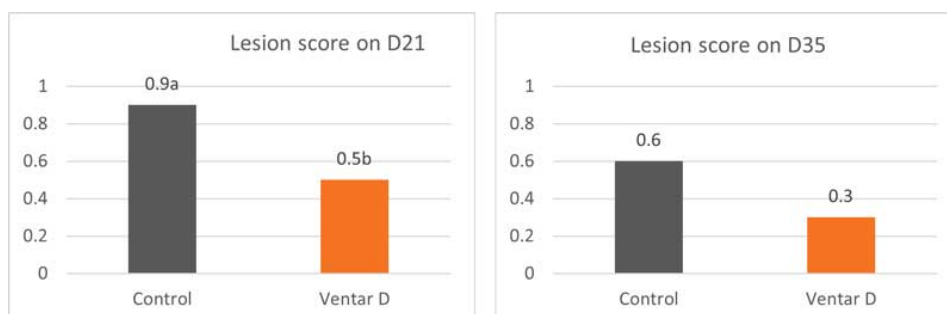
Figure 8: *Lactobacillus plantarum* exposed to 0 (left) and 1250 µg/ml (right) of Ventar D

1. In vivo-trial in poultry shows that phytomolecules reduce gut lesions

The study was conducted at Southern Poultry Feed & Research, Athens, GA (USA), over 42 days. It included in total 880 Cobb 500 broilers across 2 trial groups, with 11 repetitions per trial set-up and 40 animals per replicate floor pen. All animals received routine vaccinations at the hatchery and were healthy when starting the trial.

Control group	Built-up litter (no additive)
Ventar D group	Built-up litter + Ventar D, 100 g/MT of feed

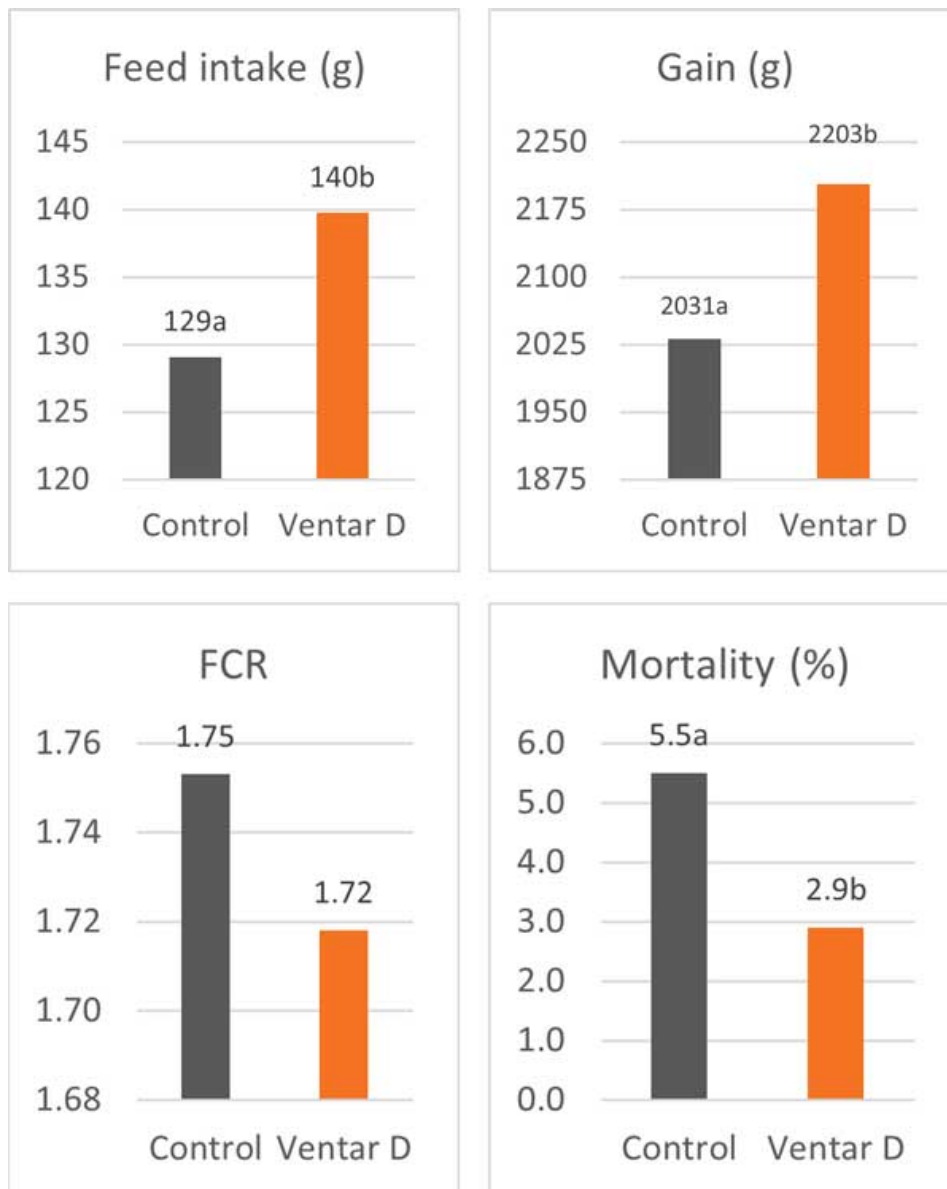
All birds received standard feed, fed as crumbles/pellets ad libitum. Feed intake by pen was recorded per feeding phase for starter (D21), grower (D35), and finisher feed (D42). Bird weights were recorded at study initiation, on D21, D35, and D42. On D21 and D35, three birds per pen were sacrificed. The GIT was scored for necrotic enteritis lesions; figures 9 and 10 show the results.



Figures 9 and 10: Lesion score on days 21 and 35

Already on day 21, the birds of the Ventar D group showed a less impacted gut mucosa, indicated by a lower lesion score. Lesions were reduced in both groups until day 35; however, the value of the Ventar D group was still better.

A less impacted gut has a higher digestion and absorption capacity, which results in better performance (FCR and weight gain) and lower mortality (figures 11-14).



Figures 11-14: Performance data of a control group compared with birds supplemented with Ventar D

The two trials show that Ventar D allows the poultry producer to proactively strengthen broilers' gut health by controlling *Clostridia perfringens* and promoting/saving beneficial bacteria such as lactobacilli. The effects of the reduction of *Clostridia* can be seen in vivo in a lower lesion score and better performance.

Toxin binders adsorb bacterial and mycotoxins

These binders have two modes of action:

They bind mycotoxins and, therefore, reduce or prevent damage to the intestinal wall so that the preconditions for *Clostridium* spp. proliferation are not generated.

Additionally, binding toxins produced by *Clostridium perfringens* can reduce the occurrence or severity of lesions: Alpha-toxin, phospholipase C, hydrolyses membrane phospholipids and damages erythrocytes, leucocytes, myocytes, and endothelial cells and causes their lipolysis (Songer, 1996), leading to necrosis and tissue damage.

Binding NetB toxin, the key virulence factor, could reduce the severity of Necrotic Enteritis.

A trial was conducted in a laboratory in Valladolid/Spain to show the high binding capacity of Solis Plus 2.0. All tests were carried out as duplicates and using a standard liquid chromatography/mass spectrometry (LC/MS/MS) quantification. Interpretation and data analysis were carried out with the corresponding software. Toxin concentrations, anti-mycotoxin agent application rates, and pH levels were set as follows:

Mycotoxin	Challenge Level	Challenge (ppb)	Solis Plus 2.0 inclusion rate	Assay time
Aflatoxin	Low	150	0.2%	30 min.
	High	1500		
Fumonisin	Low	500		
	High	5000		
Ochratoxin	Low	150		
	High	1500		

The results are shown in figure 15:

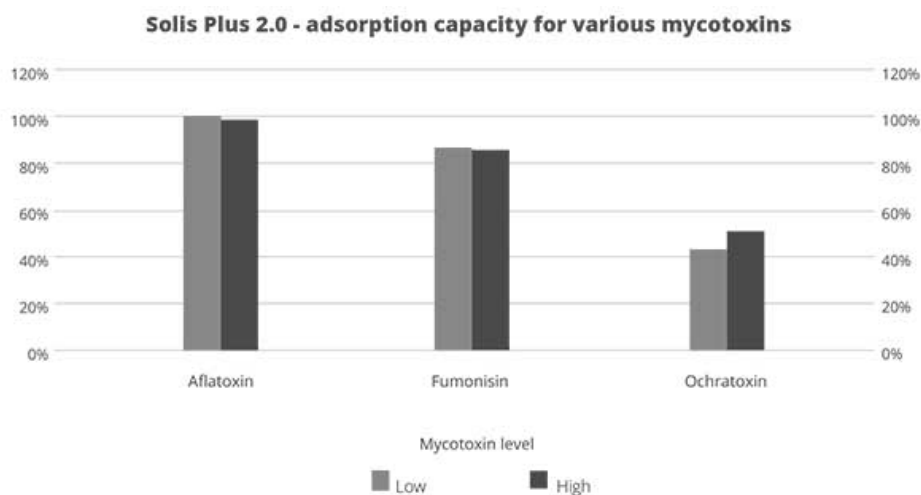


Figure 15: Adsorption capacity of Solis Plus for relevant mycotoxins

Under acidic conditions (pH 3), Solis Plus 2.0 effectively adsorbs the three tested mycotoxins at low and high contamination levels:

- Aflatoxin: 150 ppb -100 %; 1500 ppb – 98 %
- Fumonisin: 500 ppb – 87%; 5000 ppb – 86 %
- Ochratoxin: 150 ppb – more than 43 %; 1500 ppb – 52 %.

By binding harmful toxins and preventing their negative impact on the gut, toxin binders can also be a tool to reduce necrotic enteritis.

NE can be controlled - even in an antibiotic-free era

The ever-growing trend of reduced antibiotic and ionophore use increases the incidence of Necrotic Enteritis in poultry production. Especially the subclinical form, which generally goes unnoticed, results in poor feed efficiency and is a major cause of financial losses to poultry producers.

Maintaining optimum gut health is key to preventing the occurrence of Necrotic Enteritis. In the era of antibiotic-free poultry production, alternatives acting against the pathogenic bacterium and also against its predisposing factors must be considered to control this devastating disease. The industry already provides solutions like phytomolecules-based products or toxin binders to support the animals.

References:

Annett, C.B., J. R. Viste, M. Chirino-Trejo, H. L. Classen, D. M. Middleton, and E. Simko. "Necrotic enteritis: effect of barley, wheat and corn diets on proliferation of *Clostridium perfringens* type A." *Avian Pathology* 31 (2002): 599– 602. <https://doi.org/10.1080/0307945021000024544>

Antonissen G, F. Van Immerseel, F. Pasmans, R. Ducatelle, F. Haesebrouck, L. Timbermont, M. Verlinden, G.P.J.

- Janssens, V. Eeckhaut, M. Eeckhout, S. De Saeger, S. Hessenberger, A. Martel, and S. Croubels. "The mycotoxin deoxynivalenol predisposes for the development of *Clostridium perfringens*-Induced necrotic enteritis in broiler chickens. *PLoS ONE* 9 no. 9 (2014): e108775. <https://doi.org/10.1371/journal.pone.0108775>
- Antonissen, G., V. Eeckhaut, K. Van Driessche, L. Onrust, F. Haesebrouck, R. Ducatelle, R.J. Moore, and F. Van Immerseel. "Microbial Shifts Associated With Necrotic Enteritis." *Avian Pathol.* 45 no. 3 (2016): 308-312. <https://doi.org/10.1080/03079457.2016.1152625>
- Branton, S.L., F.N. Reece, and W.M. Hagler. "Influence of a wheat diet on mortality of broiler chickens associated with necrotic enteritis." *Poultry Sci.* 66 (1987): 1326-1330. <https://doi.org/10.3382/ps.0661326>
- Cejas, E., S. Pinto, F. Prosdócimo, M. Batalle, H. Barrios, G. Tellez, and M. De Franceschi. "Evaluation of quebracho red wood (*Schinopsis lorentzii*) polyphenols vegetable extract for the reduction of coccidiosis in broiler chicks." *International Journal of Poultry Science* 10 no. 5 (2011): 344-349. <https://doi.org/10.3923/ijps.2011.344.349>
- Collier, C.T., C.L. Hofacre, A.M. Payne, D.B. Anderson, P. Kaiser, R.I. Mackie, and H.R. Gaskins. "Coccidia-induced mucogenesis promotes the onset of necrotic enteritis by supporting *Clostridium perfringens* growth." *Veterinary Immunology and Immunopathology* 122 (2008):104-115. <https://doi.org/10.1016/j.vetimm.2007.10.014>
https://www.academia.edu/12692646/Coccidia-induced_mucogenesis_promotes_the_onset_of_necrotic_enteritis_by_supporting_Clostridium_perfringens_growth
- Dahiya, J.P., D. Hoehler, A.G. Van Kessel, and M.D. Drew. "Effect of different dietary methionine sources on intestinal microbial populations in broiler chickens." *Poultry Science* 86 (2007):2358-2366 <https://doi.org/10.3382/ps.2007-00133>
- Dahiya, J.P., D. Hoehler, D.C. Wilkie, A.G. van Kessel, and M.D. Drew. "Dietary glycine concentration affects intestinal *Clostridium perfringens* and *Lactobacilli* populations in broiler chickens." *Poultry Science* 84 no.12 (2005):1875-85. <https://doi.org/10.1093/ps/84.12.1875>
- Diaz Carrasco, J.M., L.M. Redondo, E.A. Redondo, J.E. Dominguez, A.P. Chacana, and M.E. Fernandez Miyakawa. "Use of plant extracts as an effective manner to control *Clostridium perfringens* induced necrotic enteritis in poultry." *BioMed Research International* (2016): Article ID 3278359. <https://dx.doi.org/10.1155/2016/3278359>
- Ducatelle, R. and F. van Immerseel. "Necrotic enteritis: emerging problem in broilers." *WATTAgNet.com - Poultry Health and Disease* (April 9, 2010). <https://www.wattagnet.com/articles/5523-necrotic-enteritis-emerging-problem-in-broilers>
- Elizondo, A.M., E.C. Mercado, B.C. Rabinovitz, and M.E. Fernandez-Miyakawa. "Effect of tannins on the in vitro growth of *Clostridium perfringens*." *Veterinary Microbiology* 145 no. 3-4 (2010): 308-314. <https://doi.org/10.1016/j.vetmic.2010.04.003>
- Engberg, R.M., M.S. Hedemann, and B.B. Jensen. "The influence of grinding and pelleting of feed on the microbial composition and activity in the digestive tract of broiler chickens." *British Poultry Science* 43 no. 4 (2002):569-579. <https://doi.org/10.1080/0007166022000004480>
- Fischetti, V.A. "Bacteriophage endolysins: A novel anti-infective to control Gram-positive pathogens." *J Med Microbiol.* 300 no. 6 (2010): 357-362. <https://doi.org/10.1016/j.jimm.2010.04.002>
- Gillor, O., A. Etzion and M.A. Riley. "The dual role of bacteriocins as anti- and probiotics." *Appl Microbiol Biotechnol.* 81 no. 4 (2008): 591-606. <https://doi.org/10.1007/s00253-008-1726-5>
- Hofacre, C.L., J.A. Smith, and G.F. Mathis. "Invited Review. An optimist's view on limiting necrotic enteritis and maintaining broiler gut health and performance in today's marketing, food safety, and regulatory climate." *Poultry Science* 97 (2018):1929-1933. <https://dx.doi.org/10.3382/ps/pey082>
- Jung, S.J., R. Houde, B. Baurhoo, X. Zhao, and B. H. Lee. "Effects of galacto-oligosaccharides and a bifidobacteria lactis-based probiotic strain on the growth performance and fecal microflora of broiler chickens." *Poultry Science* 87 (2008):1694-1699. <https://doi.org/10.3382/ps.2007-00489>

Kaldhusdal and Skjerve. "Association between cereal contents in the diet and incidence of necrotic enteritis in broiler chickens in Norway." *Preventive Veterinary Medicine* 28 (1996):1-16.

[https://doi.org/10.1016/0167-5877\(96\)01021-5](https://doi.org/10.1016/0167-5877(96)01021-5)

https://www.academia.edu/17521917/Association_between_cereal_contents_in_the_diet_and_incidence_of_necrotic_enteritis_in_broiler_chickens_in_Norway

Keyburn, A. L., S. A. Sheedy, M. E. Ford, M. M. Williamson, M. M. Awad, J. I. Rood, and R. J. Moore. "Alpha-toxin of *Clostridium perfringens* is not an essential virulence factor in necrotic enteritis in chickens." *Infect. Immun.* 74 (2006): 6496-6500. <https://doi.org/10.1128/IAI.00806-06>

Keyburn, A.L., J.D. Boyce, P. Vaz, T.L. Bannam, M.E. Ford, D. Parker, A. Di Rubbo, J.I. Rood, and R.J. Moore. "NetB, a new toxin that is associated with avian necrotic enteritis caused by *Clostridium perfringens*." *PLoS Pathog* 4 no. 2, e26 (2008): 0001-0011. <https://doi.org/10.1371/journal.ppat.0040026>

Kim, G.-B., Y. M. Seo , C. H. Kim , and I. K. Paik. "Effect of dietary prebiotic supplementation on the performance, intestinal microflora, and immune response of broilers." *Poultry Science* 90 (2011):75-82.

<https://doi.org/10.3382/ps.2010-00732>

Knap, I., B. Lund, A. B. Kehlet, C. Hofacre, and G. Mathis. "Bacillus licheniformis prevents necrotic enteritis in broiler chickens." *Avian Diseases* 54 no. 2 (2010):931-935. <https://doi.org/10.1637/9106-101509-ResNote.1>

Knarreborg, A., M.A. Simon, R.M. Engberg, B.B. Jensen, and G.W. Tannock. "Effects of Dietary Fat Source and Subtherapeutic Levels of Antibiotic on the Bacterial Community in the Ileum of Broiler Chickens at Various Ages." *Applied and Environmental Microbiology* 68 no. 12 (2002): 5918-5924.

<https://doi.org/10.1128/AEM.68.12.5918-5924.2002>

Kocher, A. and M. Choct. "Improving broiler chicken performance. The efficacy of organic acids, prebiotics and enzymes in controlling necrotic enteritis." *Australian Government-Rural Industries Research and Development Corporation*. Publ. no. 08/149 (2008).

<https://www.agrifutures.com.au/wp-content/uploads/publications/08-149.pdf>

Kondo, F. "In vitro lecithinase activity and sensitivity to 22 antimicrobial agents of *Clostridium perfringens* isolated from necrotic enteritis of broiler chickens." *Research in veterinary Science* 45 (1988): 337-340.

[https://doi.org/10.1016/S0034-5288\(18\)30961-5](https://doi.org/10.1016/S0034-5288(18)30961-5)

Kubena, L.F., J.A. Byrd, C.R. Young, and D.E. Corrier. "Effects of tannic acid on cecal volatile fatty acids and susceptibility to *Salmonella typhimurium* colonization in broiler chicks." *Poultry Science* 80, no. 9, pp. 1293-1298, 2001.

<https://doi.org/10.1093/ps/80.9.1293>

M'Sadeq S.A., Shubiao Wu, Robert A. Swick, Mingan Choct. "Towards the control of necrotic enteritis in broiler chickens with in-feed antibiotics phasing-out worldwide." *Animal Nutrition* 1 (2015): 1-11.

<https://dx.doi.org/10.1016/j.aninu.2015.02.004>

Mathipa, M.G. and M.S. Thantsha. "Probiotic engineering: towards development of robust probiotic strains with enhanced functional properties and for targeted control of enteric pathogens." *Gut Pathog.* 9 no. 28 (2017).

<https://doi.org/10.1186/s13099-017-0178-9>

McDevitt, R.M., J.D. Brooker, T. Acamovic, and N.H.C. Sparks. "Necrotic enteritis, a continuing challenge for the poultry industry." *World's Poultry Science Journal* 62; World's Poultry Science Association (June 2006).

<https://doi.org/10.1079/WPS200593>

Miller, R.W., J. Skinner, A. Sulakvelidze, G.F. Mathis, and C.L. Hofacre. "Bacteriophage therapy for control of Necrotic Enteritis of broiler chickens experimentally infected with *Clostridium perfringens*." *Avian Diseases* 54 no. 1 (2010): 33-40. <https://doi.org/10.1637/8953-060509-Reg.1>

Mitsch, P., K. Zitterl-Eglseer, B. Köhler, C. Gabler, R. Losa, and I. Zimpernik. "The Effect of Two Different Blends of Essential Oil Components on the Proliferation of *Clostridium perfringens* in the Intestines of Broiler Chickens." *Poultry Science* 83 (2004):669-675. <https://doi.org/10.1093/ps/83.4.669>

Mitchell, A. "Choosing the right coccidiosis vaccine for layer and breeder chickens." *The Poultry Site* March 21 (2017). <https://thepoultrysite.com/articles/choosing-the-right-coccidiosis-vaccine-for-layer-and-breeder-chickens>

- Olkowski, A.A., C. Wojnarowicz, M. Chirino-Trejo, B. Laarveld, and G. Sawicki. "Sub-clinical necrotic enteritis in broiler chickens: Novel etiological consideration based on ultra-structural and molecular changes in the intestinal tissue." *Veterinary Science* 85 (2008): 543-553. <https://doi.org/10.1016/j.rvsc.2008.02.007>
- Pan, D. and Z. Yu. "Intestinal microbiome of poultry and its interaction with host and diet." *Gut Microbes* 5 no. 1 (2014): 108-119. <https://dx.doi.org/10.4161/gmic.26945>
- Robert Koch Institut. "Grundwissen Antibiotikaresistenz". https://www.rki.de/DE/Content/Infekt/Antibiotikaresistenz/Grundwissen/Grundwissen_inhalt.html#:~:text=Wenn%20ein%20neues%20Antibiotikum%20auf,%C3%BCberleben%20und%20vermehren%20sich%20weiter.
- Rougière, N. and B. Carré. "Comparison of gastrointestinal transit times between chickens from D + and D- genetic lines selected for divergent digestion efficiency." *Animal* 4 no. 11 (2010): 1861-1872. <https://doi.org/10.1017/S1751731110001266>
- Santos, F.B.O., B.W. Sheldon, A.A. Santos Jr., and P.R. Ferket. "Influence of housing system, grain type, and particle size on Salmonella colonization and shedding of broilers fed triticales or corn-soybean meal diets." *Poultry Science* 87 (2008): 405-420. <https://dx.doi.org/10.3382/ps.2006-00417>
- Schiavone, A. , K. Guo, S. Tassone, L. Gasco, E. Hernandez, R. Denti, and I. Zoccarato. "Effects of a Natural Extract of Chestnut Wood on Digestibility, Performance Traits, and Nitrogen Balance of Broiler Chicks." *Poult Sci.* 87 no. 3 (2008): 521-527. <https://doi.org/10.3382/ps.2007-00113>
- Singh, Y., V. Ravindran, T.J. Wester, A.L. Molan, and G. Ravindran. "Influence of feeding coarse corn on performance, nutrient utilization, digestive tract measurements, carcass characteristics, and cecal microflora counts of broilers." *Poultry Science* 93 (2014): 607-616. <https://dx.doi.org/10.3382/ps.2013-03542>
- Skrivanova, E., M. Marounek, V. Benda, and P. Brezina. "Susceptibility of Escherichia coli, Salmonella sp. and Clostridium perfringens to organic acids and monolaurin." *Veterinarni Medicina* 51 no. 3 (2006): 81-88. <https://doi.org/10.17221/5524-VETMED>
- Songer, J.G. "Clostridial Enteric Diseases of Domestic Animals." *Clinical Microbiology Reviews* 9 no. 2 (1996): 216-234. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC172891/pdf/090216.pdf>
- Stanley D., Wu S.-B., Rodgers N., Swick R.A., and Moore R.J. "Differential Responses of Cecal Microbiota to Fishmeal, Eimeria and Clostridium perfringens in a Necrotic Enteritis Challenge Model in Chickens." *PLoS ONE* 9 no. 8 (2014): e104739. <https://doi.org/10.1371/journal.pone.0104739>
- Tan, L., D. Rong, Y. Yang, and B. Zhang. "Effect of Oxidized Soybean Oils on Oxidative Status and Intestinal Barrier Function in Broiler Chickens." *Brazilian Journal of Poultry Science* 18 no. 2 (2018): 333-342. <https://dx.doi.org/10.1590/1806-9061-2017-0610>
- Tan, L., D. Rong, Y. Yang, and B. Zhang. "The Effect of Oxidized Fish Oils on Growth Performance, Oxidative Status, and Intestinal Barrier Function in Broiler Chickens." *J. Appl. Poult. Res.* 28 (2019): 31-41. <http://dx.doi.org/10.3382/japr/pfy013>
- ThePoultrySite. "Necrotic Enteritis. Disease Guide". <https://thepoultrysite.com/disease-guide/necrotic-enteritis>
- Timbermont L., A. Lanckriet, J. Dewulf, N. Nollet, K. Schwarzer, F. Haesebrouck, R. Ducatelle, and F. Van Immerseel. "Control of Clostridium perfringens-induced necrotic enteritis in broilers by target-released butyric acid, fatty acids and essential oils." *Avian Pathol.* 39 no. 2 (2010): 117-21. <https://doi.org/10.1080/03079451003610586>
- Tsiouris, V. "Poultry management: a useful tool for the control of necrotic enteritis in poultry." *Avian Pathol.* 45 no. 3 (2016):323-325. <https://doi.org/10.1080/03079457.2016.1154502>
- Van der Most, P.J., B. de Jong, H.K. Parmentier and S. Verhulst. "Trade-off between growth and immune function: a meta-analysis of selection experiments." *Functional Ecology* 25 (2011): 74-80. <https://doi.org/10.1111/j.1365-2435.2010.01800.x>
- Van der Sluis, W. "Clostridial enteritis is an often underestimated problem." *Worlds Poult. Sci. J.* 16 (2000):42-43.
- Van der Suis, W. "Necrotic enteritis kills birds and profits." *Poultry World* Apr5 (2013).

<https://www.poultryworld.net/Health/Articles/2013/4/Necrotic-enteritis-kills-birds-and-profits-1220877W/>

Van Immerseel, F., J. De Buck, F. Pasmans, G. Huyghebaert, F. Haesebrouck, and R. Ducatelle. "Clostridium perfringens in poultry: an emerging threat of animal and public health." *Avian Pathology* 33 (2004): 537-549. <https://doi.org/10.1080/03079450400013162>

Van Immerseel, F., J.I. Rood, R.J. Moore, and R.W. Titball. "Rethinking our understanding of the pathogenesis of necrotic enteritis in chickens." *Trends in Microbiology* 17 no. 1 (2008):32-36. <https://doi.org/10.1016/j.tim.2008.09.005>

Wade, B., A. Keyburn. "The true cost of necrotic enteritis." *World Poultry* 31 no. 7 (2015): 16-17. <https://www.poultryworld.net/Meat/Articles/2015/10/The-true-cost-of-necrotic-enteritis-2699819W/>

Wade, B., A.L. Keyburn, T. Seemann, J.I. Rood, and R.J. Moore. "Binding of Clostridium perfringens to collagen correlates with the ability to cause necrotic enteritis in chickens." *Veterinary Microbiology* 180 no. 3-4 (2015): 299-303. <https://doi.org/10.1016/j.vetmic.2015.09.019>

Williams, R.B. "Intercurrent coccidiosis and necrotic enteritis of chickens: rational, integrated disease management by maintenance of gut integrity." *Avian Pathology* 34 no. 3 (2005):159-180. <https://doi.org/10.1080/03079450500112195>

Yang , C.M., G.T. Cao, P.R. Ferket, T.T. Liu, L. Zhou, L. Zhang, Y.P. Xiao, and A. G. Chen. "Effects of probiotic, Clostridium butyricum, on growth performance, immune function, and cecal microflora in broiler chickens." *Poultry Science* 91 (2012): 2121-2129. <https://dx.doi.org/10.3382/ps.2011-02131>